

Creating Reliable Data – a Challenge for Non-target Screening

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Non-target screening is in daily use to manage chemicals at the International Rhine monitoring station

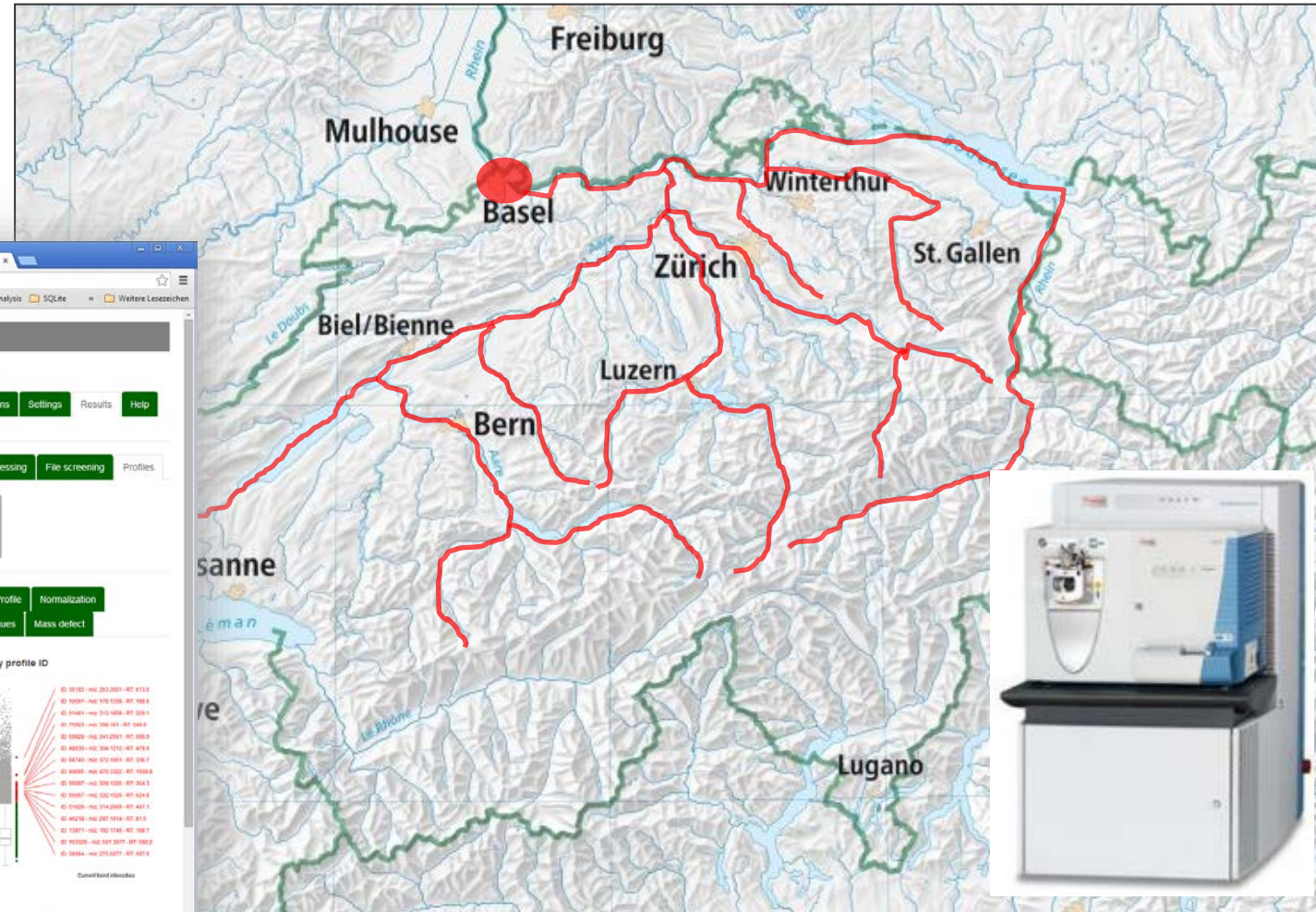


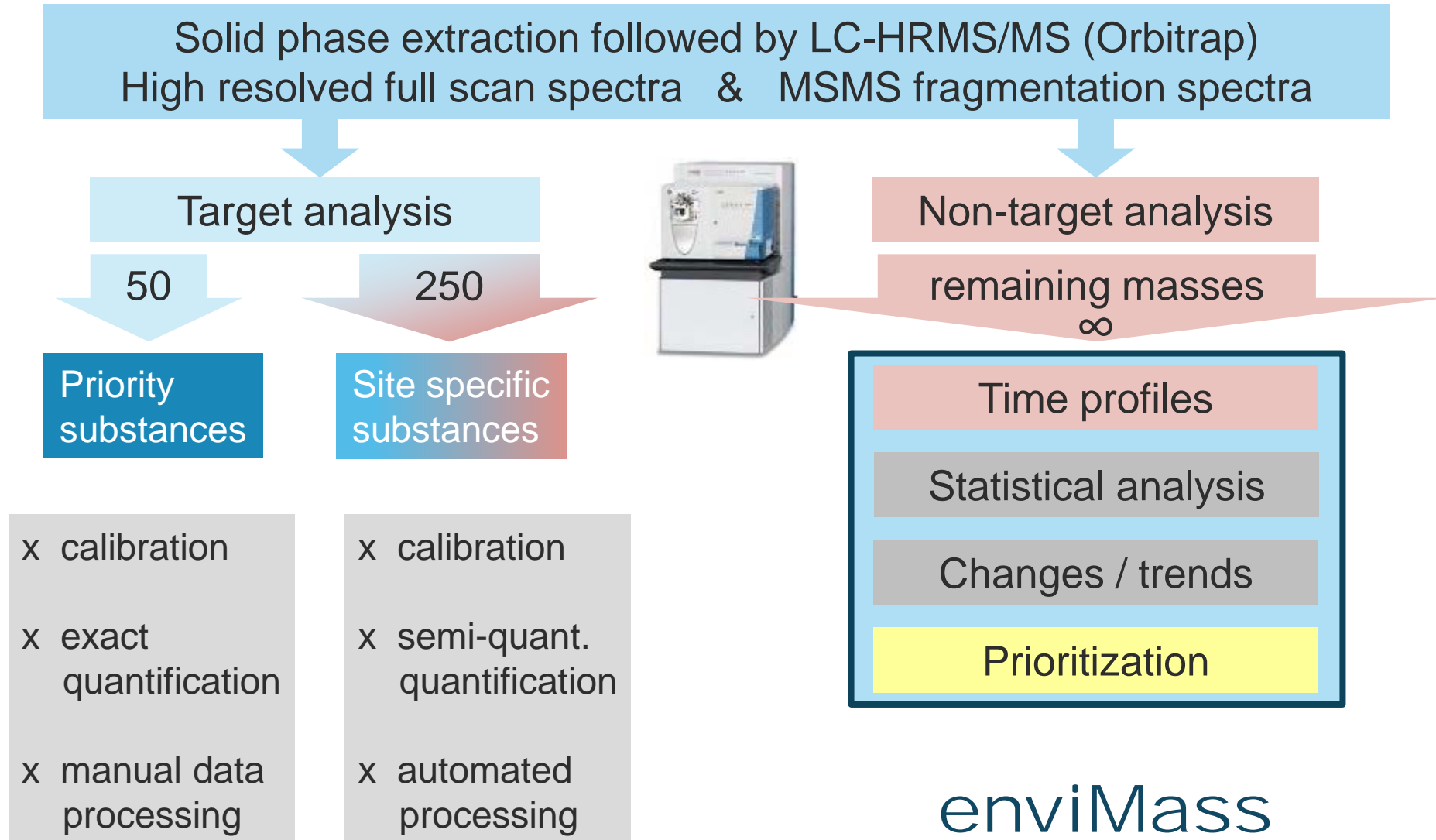
Departement für Wirtschaft, Soziales und Umwelt des Kantons Basel-Stadt

Amt für Umwelt und Energie

enviMass

Tasks	Done?
1 Data available?	TRUE
2 Peak pick?	TRUE
3 QC?	TRUE
4 Isotope pattern?	TRUE
5 m/z recal?	TRUE
6 Aligned?	FALSE
7 Intensity norm?	TRUE
8 Profiled?	TRUE
9 IS norm?	TRUE
10 Trend-Blind?	TRUE
11 IS file-screen?	FALSE
12 Target file-screen?	FALSE
13 Compon?	FALSE
14 IS comp-screen?	FALSE
15 Target comp-screen?	FALSE
16 Homologues?	FALSE
17 Mass defect?	FALSE

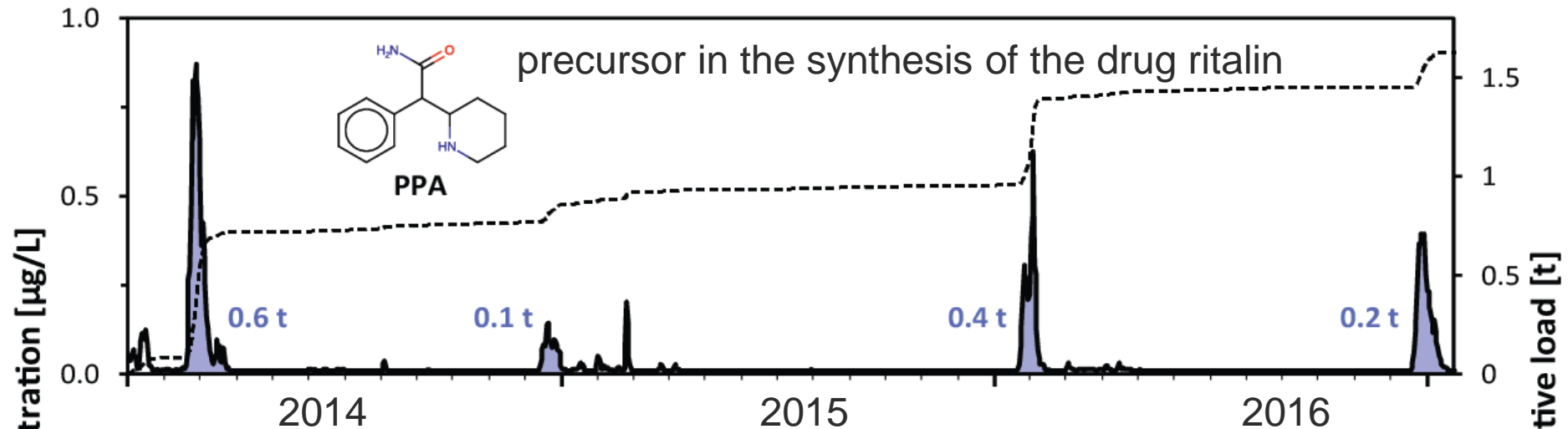






Prioritization using time profiles

Previously unknown chemicals detected due to “stand-out” patterns



Identification

- Molecular formula assignment
- Database search
- Prioritization of hits with information on industrial production
- Confirmation with reference standard

► 10 major spills of non-target compounds in 2014 with > 25 tons of load

Experiences/tools of the NORMAN network

Challenges

- Suspect lists
- Quality control
- False positives versus false negatives
- In-source fragmentation
- Identification confidence
- Semi-quantification without standard

Conclusion & points for the discussion



Example
Monitoring of Swiss groundwater



- > **70 organisations**
academia, governmental organisations,
research centers, industry

The mission of the NORMAN network is to:

- Exchange of information and collection of data on **emerging environmental substances**
- **Validation and harmonisation** of monitoring tools
- Bottom up activities
- Science to policy interface

8 working groups, one on **NTS with > 30 participating organizations**

<https://www.norman-network.net/?q=node/252>



Cross-Working Group Activity Non-target Screening (NTS)

Non-target screening techniques for environmental monitoring



Comparison & harmonization of NTS methods in Europe

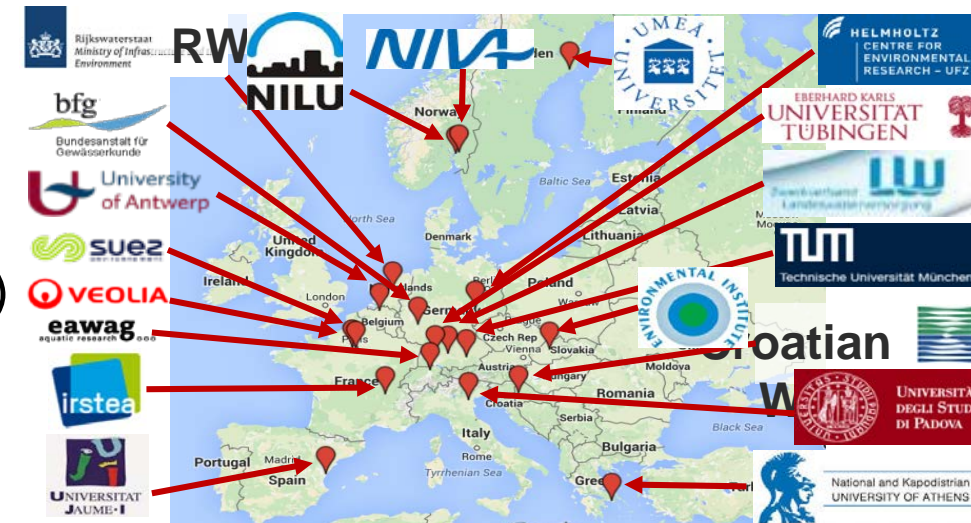
- **Collaborative trials using LC/GC-HRMS/MS:** on water (Schymanski et al. ABC, 2015) and dust (Rostkowski et al, ABC, 2019), in 2019 planned for biota and passive samplers

Databases & Tools

- Home for **MassBank**, **Norman Suspect Lists Exchange**, **Digital Sample Freezing platform** (Alygizakis et al, TrAc 2019)
- **Retention time index** (Aalizadeh, J. Chem Inf Mod 2016 & J Haz Mat 2019)
- **NormaNews exchange** (Alygizakis et al, ES&T 2018)

Communication

- NTS group meetings
- NTS workshops / training courses (e.g. CH 2014, N 2017)
- NTS workshop for regulators in Brussels, 2018

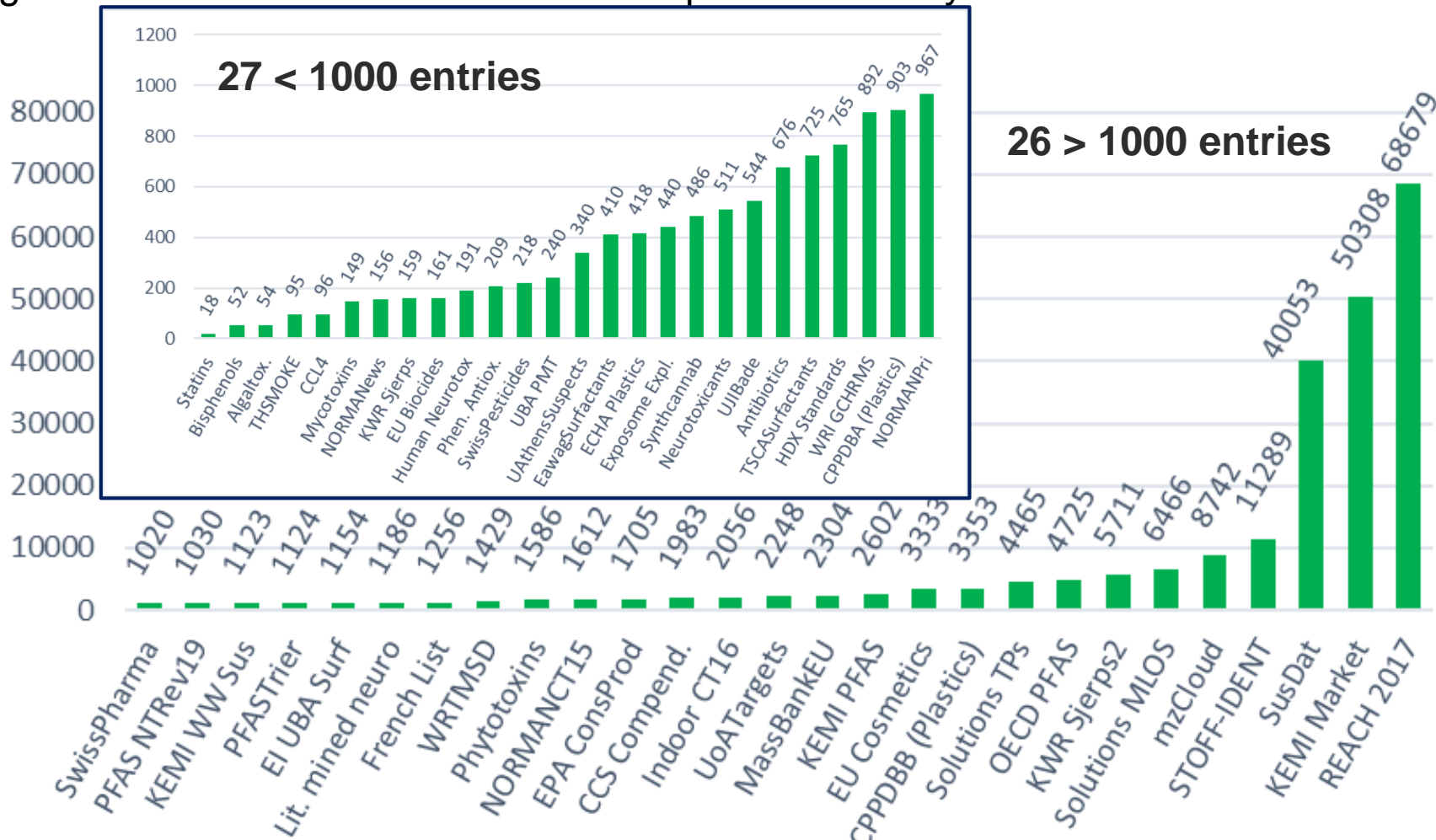


NORMAN databases: Suspect List Exchange

- <https://www.norman-network.com/?q=suspect-list-exchange>
- 53 lists available ... specialist collections to market lists
 - Integrated in NORMAN Databases & CompTox Chemistry Dashboard

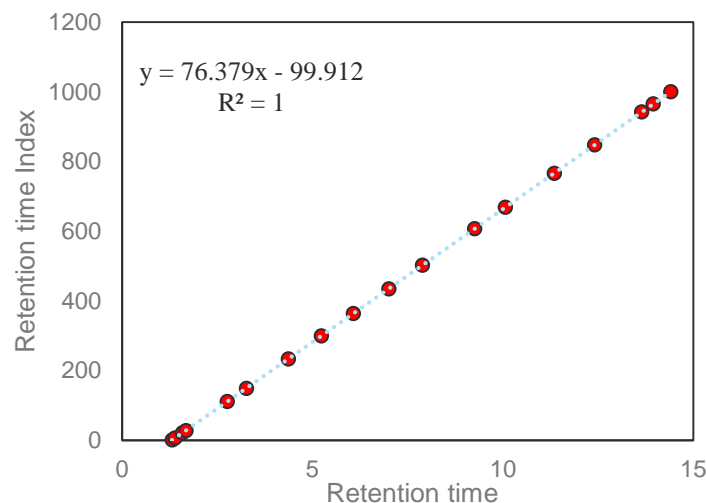


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Retention Time Indices for RPLC-(+)ESI-HRMS

Quantitative structure retention relationship models for RTI system using chemical structure information



$$RTI = 76.379(RT) - 99.912$$

Predicted by QSRR model:
C18 column, H₂O/MeOH formic acid gradient

	Training			Test		
	R2	RMSE	F	R2	RMSE	F
MLR	0.835	92.575	1515.130	0.870	83.184	426.416
SVM	0.861	84.869	1838.745	0.880	80.029	467.038

$$RTI = \frac{(RTx - RTmin)}{(RTmax - RTmin)} * 1000$$

Calibrants	RT (Acclaim C18)	RTI
Guanylurea	1.31	1.00
Amitrole	1.39	6.11
Histamine	1.58	20.63
Chlormequat	1.67	27.50
Methamidophos	2.76	110.77
Vancomycin	3.26	148.97
Cefoperazone	4.36	233.00
Trichlorfon	5.23	299.47
Butocarboxim	6.07	363.64
Dichlorvos	7	434.68
Tylosin	7.88	501.91
TCMTB	9.25	606.57
rifaximin	10.06	668.45
Spinosad A	11.34	766.23
Emamectin B1a	12.4	847.21
Avermectin B1a	13.64	941.94
Nigericin	13.94	964.86
Ivermectin B1a	14.4	1000.00

Online Platform to Calculate Experimental and Predicted Retention Time Indices

Development and Prediction of Retention Time Indices for LC-HRMS (version 2.0.0)



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Select the target ESI:

- +ESI
 -ESI

Select the RTI versus tR calibration mode:

- Prediction limits
 Auto-calibrate
 Manual

Upload RTI calibrants data...

Browse... Calibrants_pos_UOA.csv

Upload complete

Default max. file size 1MB (*.csv file)

[Click here to build the calibration curve...](#)

[Click here to restart the current session...](#)

[Trace Analysis & Mass Spectrometry Group](#)

[NORMAN Suspect list Exchange website](#)

Retention Time Indices for LC-HRMS (version 2.0.0):

[About](#) [Calibrants](#) [Single compound](#) [Batch mode](#) [Comparison of Experimental RTIs](#) [Chemical Curation](#) [NORMAN network](#) [FAQ](#)

Select the uncertainty measurement:

- OTrAMS
 Chemical space boundary

Enter the SMILES of a compound here:

Enter the tR for specific ESI mode

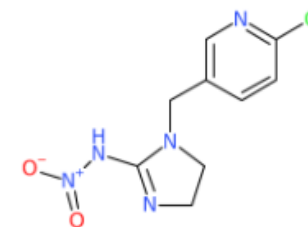
Canonical SMILES: O=[N+][O-]NC1=NCCN1CC2=CN=C(Cl)C=C2

InChIKey: YWTYJOPNNQFBPC-UHFFFAOYSA-N

Experimental tR: 4.2

The ESI selected: +ESI

Estimate RTI & its uncertainty



Status: Processed

Experimental RTI: 220.88

Predicted RTI: 250.83

Experimental tR: 4.2 min

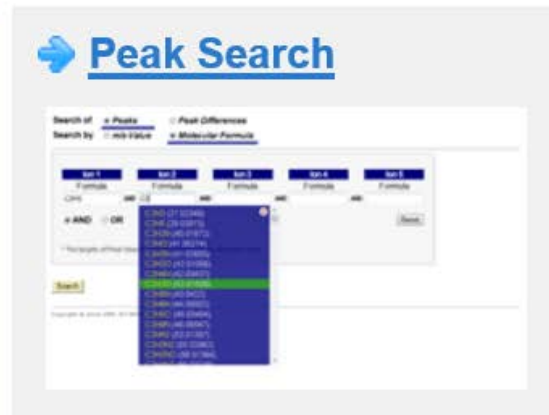
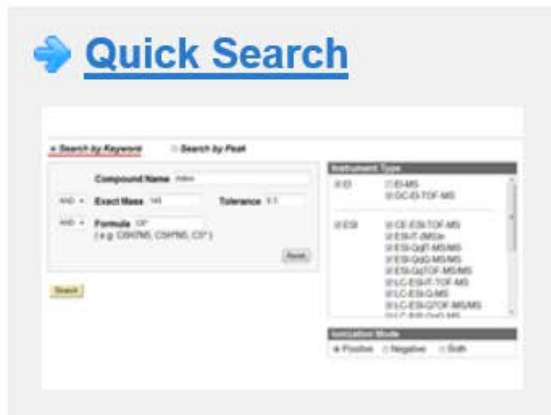
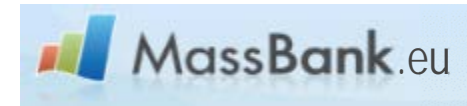
Predicted tR: 4.59 min

Uncertainty: Exp. & Pred. tR are accepted for this candidate (box1)

RTI vs tR calibration curve: $RTI = 76.3788 * (tR) - 99.9116 >>> (R^2 = 1)$

[Save the prediction results...](#)

- **European NORMAN MassBank:**
currently ~53'000 spectra of ~16'000 compounds
from 15 main instrument types and 40 institutions



- **MoNa: MassBank of North America:**
> 200'000 mass spectra including in silico spectra & European MassBank
- **mzCloud - HighChem:**
17'000 compounds, highly curated, mostly Orbitrap

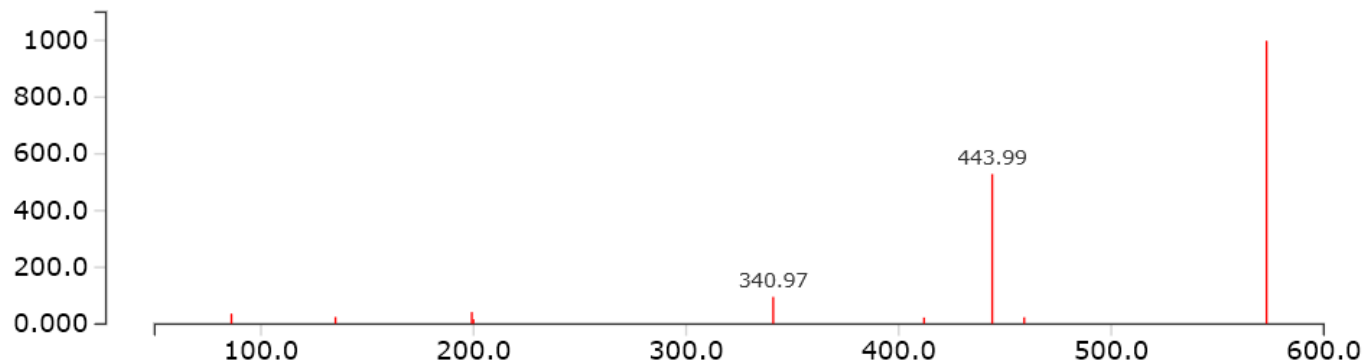


MassBank Record: ET201801

[Home](#) | [Search](#) | [Record Index](#) | [Data Privacy](#) | [Imprint](#) | MassBank ID:

PRZ_M573; LC-ESI-QFT; MS2; CE: 10; R=35000; [M+H]⁺

Mass Spectrum



ACCESSION: ET201801

RECORD_TITLE: PRZ_M573; LC-ESI-QFT; MS2; CE: 10; R=35000; [M+H]⁺

DATE: 2016.03.01

AUTHORS: , A. Roesch, E. Schymanski, J. Hollender, Department of Environmental Chemistry, Eawag

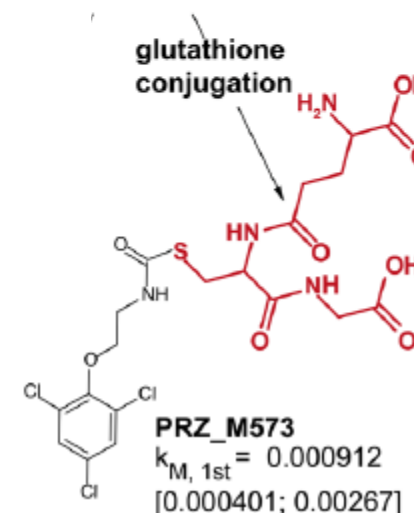
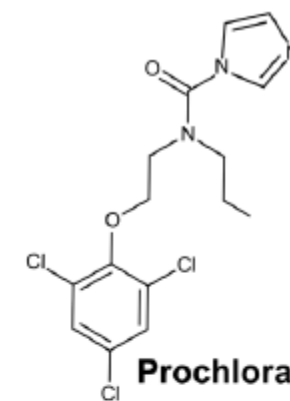
LICENSE: [CC BY](#)

COPYRIGHT: Copyright (C) 2015 Eawag, Duebendorf, Switzerland

PUBLICATION: Roesch, A.; Anliker, S.; Hollender, J. How Biotransformation Influences Toxicokinetics of Azole Fungicides in

COMMENT: CONFIDENCE Tentative identification only (Level 3)

COMMENT: INTERNAL_ID 2018

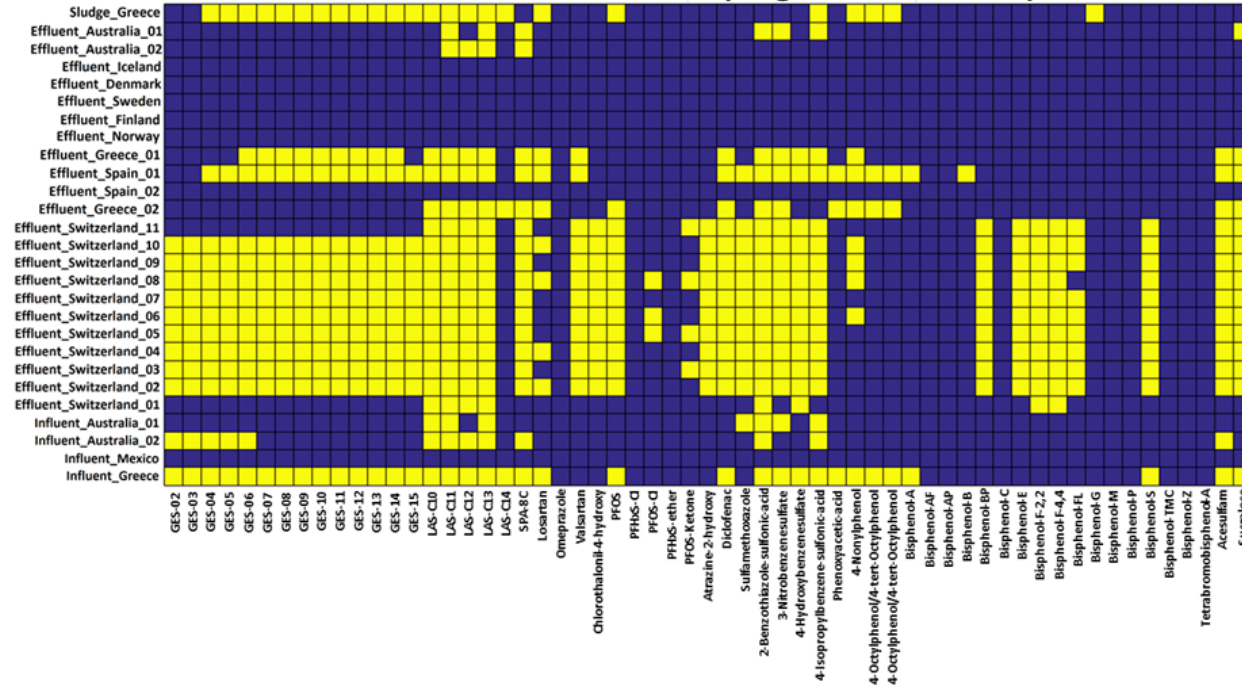


NormaNews: Sharing of emerging contaminants

Retrospective screening of emerging suspects



Wastewater matrices (Negative Ionization)



Dashboard
NORMANEWS

- 150 suspect compounds
- HRMS data from 14 countries
- QA/QC

14-hydroxy-3,6,9,12-tetraoxatetradecyl...
NOCAS_881042

1H-Benzotriazole-5-carboxylic acid
23814-12-2

3,3'-Dimethylbisphenol A
79-97-0

4-(Dodecan-6-yl)benzene-1-sulfonic acid
23003-92-1

Digital Sample Freezing Platform – DSFP

- A digital specimen bank of georeferenced HRMS data

NORMAN Digital Sample Freezing Platform

Results NORMAN-ECHA

Screening of REACH compounds in samples from the Black Sea

Selection of data

Compounds with:

>=4 fragments

Apply changes

Appearance

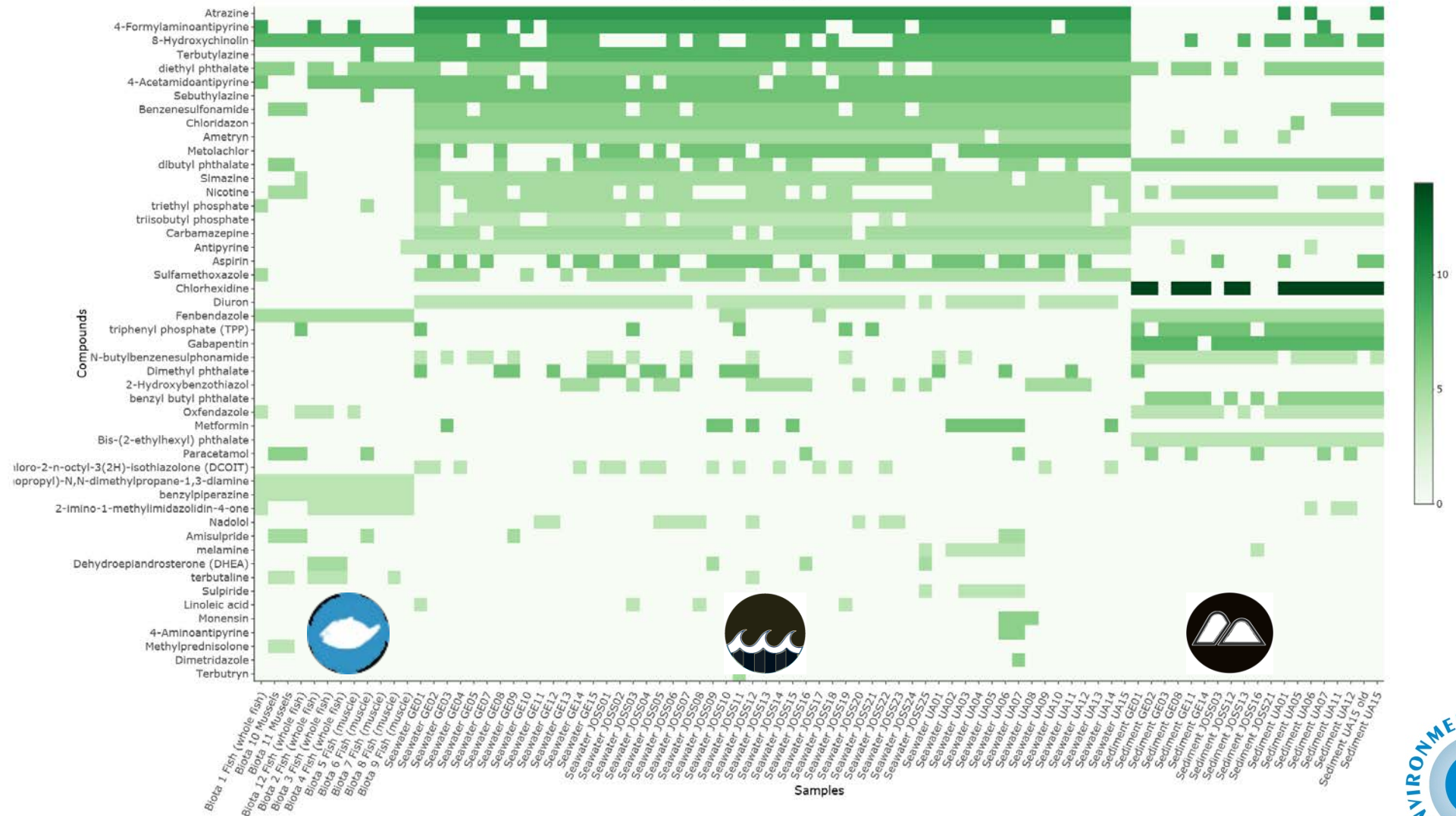
Height: 330

Width: 300

Size Letters Y: 10

Size Letters X: 10

Rotation of labels on x-axis: 65



Interactive heatmap available at <http://norman-data.eu/NORMAN-REACH>, Algyzakis et al., TrAC 2019

How can non-target screening techniques support environmental monitoring and chemicals management?

- NTS can improve the **identification of problematic substances** and **support regulatory processes in environmental and chemical legislation** (e.g. WFD, the Marine Strategy Framework Directive, REACH)
- NTS can be a **first screening step in the exposure assessment chain** but **does not replace target monitoring**.
- **Harmonized NTS protocols** and **minimum quality requirements** should be established.
- **New protocols / infrastructures** are needed for efficient NTS data management, evaluation and sharing.
- **Training** would be beneficial to make NTS more widely accessible.
- **Synergies** between NTS and effect-based methods should be strengthened



- > 80 participants from
 - Ministries, agencies
 - Regulatory institutes
 - Research institutes
 - Industry
 - Academics

responsible for

- Environmental/chemical legislation
- Food/drinking water safety
- Human biomonitoring

- **Key input:**
 - **experience of NORMAN members** from trials etc.
 - Different approaches for different regulatory questions where NTS can help (monitoring, prioritization,...)
 - Guidance of German Water Chemistry Society on Suspect and Non-target screening in water analysis, draft
 - Important topics: **Number & type of blanks, Replicate samples/ replicate analyses, Compound domains covered vs. LC/GC & MS method setup, data dependent vs. data independent acquisition**
- Presentation of draft version at the SWEMSA 2019 workshop (TU Munich, 21-23/10/19)
- Discussion on general assembly end of November in Milano
- Publish in 2020 with open access in a peer-reviewed journal

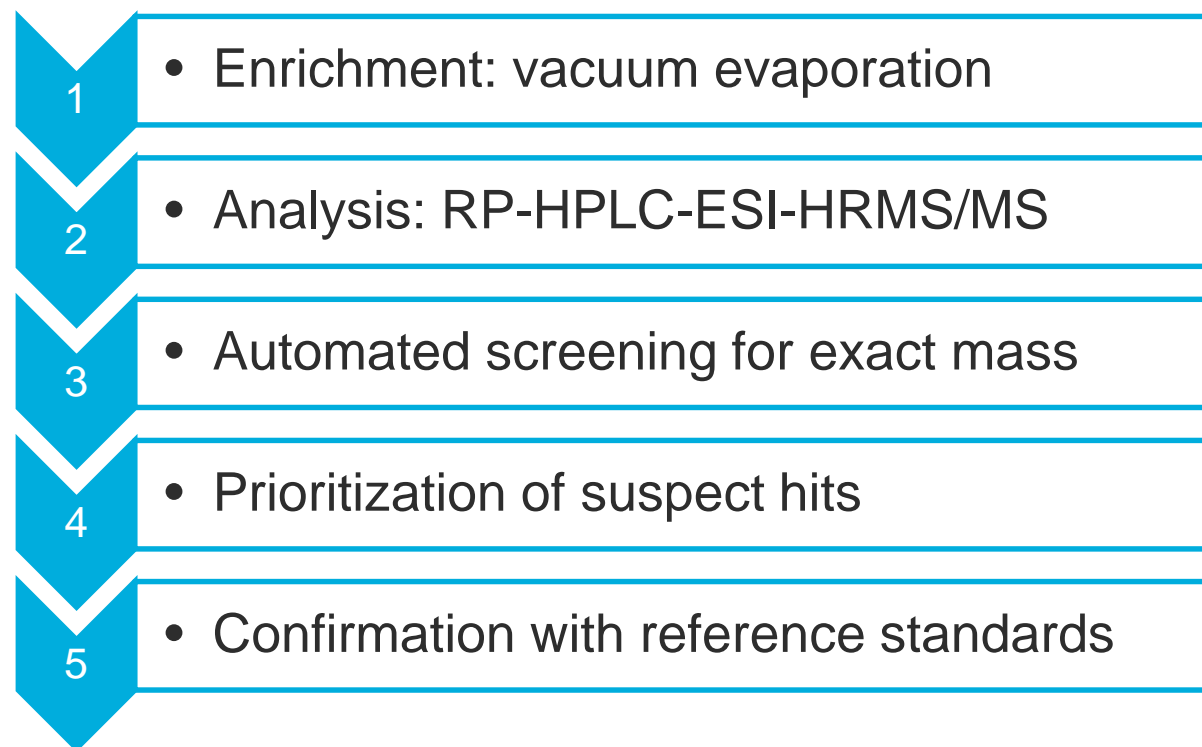


Suspect screening of overlooked pesticide TPs in Swiss groundwater

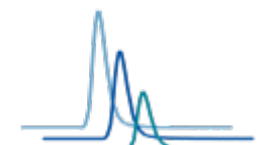
31 groundwater wells with intensive agriculture

(Swiss national monitoring sites)

Workflow

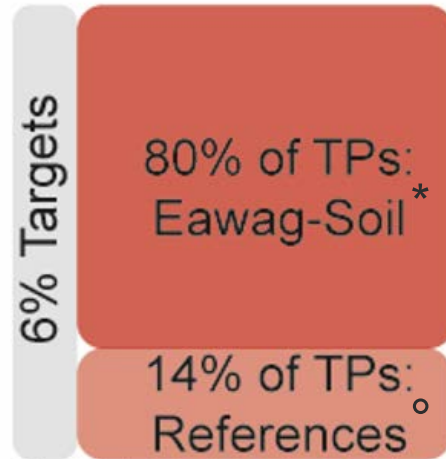


QExactivePlus
R= 140000



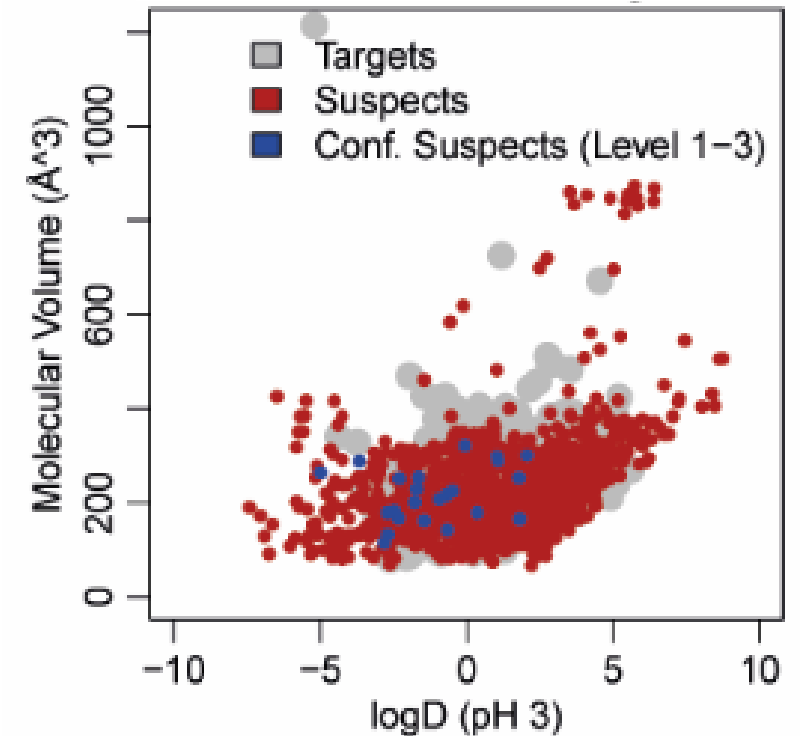
Smart Suspect Screening of pesticide transformation products in groundwater

1120 TPs
of ca. 300 pesticides



*~1000 pesticide TPs, **enviPath**
from European pesticide registration
(Latino et al. ESPI 2017)
° i.a. PPBD, Lewis et al. HERA 2016,
Reemtsma et al., Wat Res 2013

LC-ESI-MS amenability

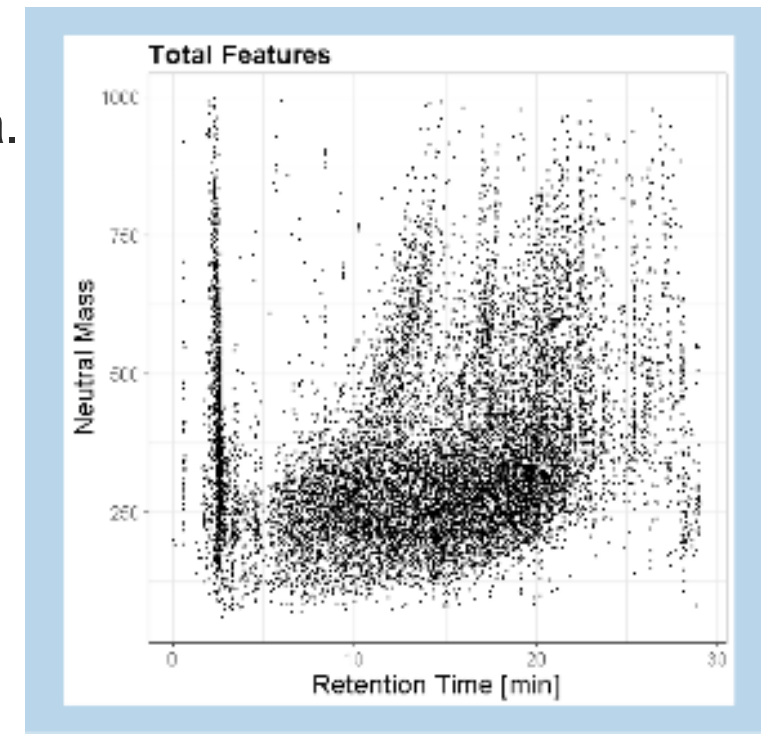


95%: similar polarity as targets
99% contain N, O, S atom

- Appropriate suspect list
- Appropriate method for compounds on suspect list

Quality control is necessary for **instrumental analysis**, but also **data processing!**

- Perform **QC samples** in each batch of samples (e.g., composite samples of each matrix, spiked samples, standard mixtures) and a sufficient number of sample processing and instrument blanks
- Use **internal standards** (spiked standards) for QC of analytical performance, but also of peak detection, sample alignment, mass accuracy.
- Take the time to adjust the **peak detection settings to your data**. Automatic parameter optimization is available in some workflows (e.g. XCMS, EnviMass).



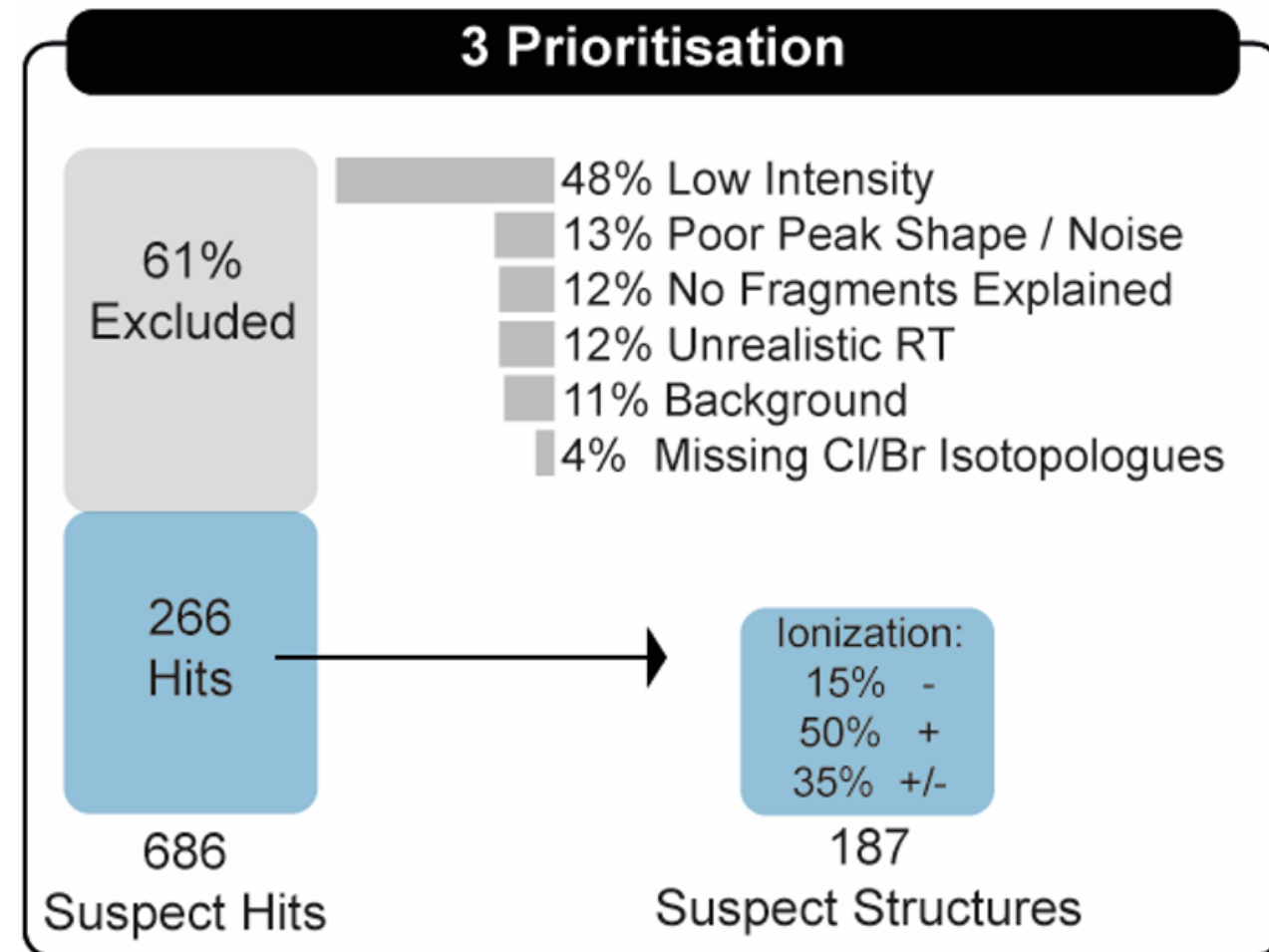
Quality control: False positives versus false negatives

Example: aim to not overlook TPs with low ionization efficiency

Validation of workflow

(peak picking, Rt alignment, isotopologues grouping, background subtraction, suspect screening, CD 2.1 Thermo) **with >200 internal standards**

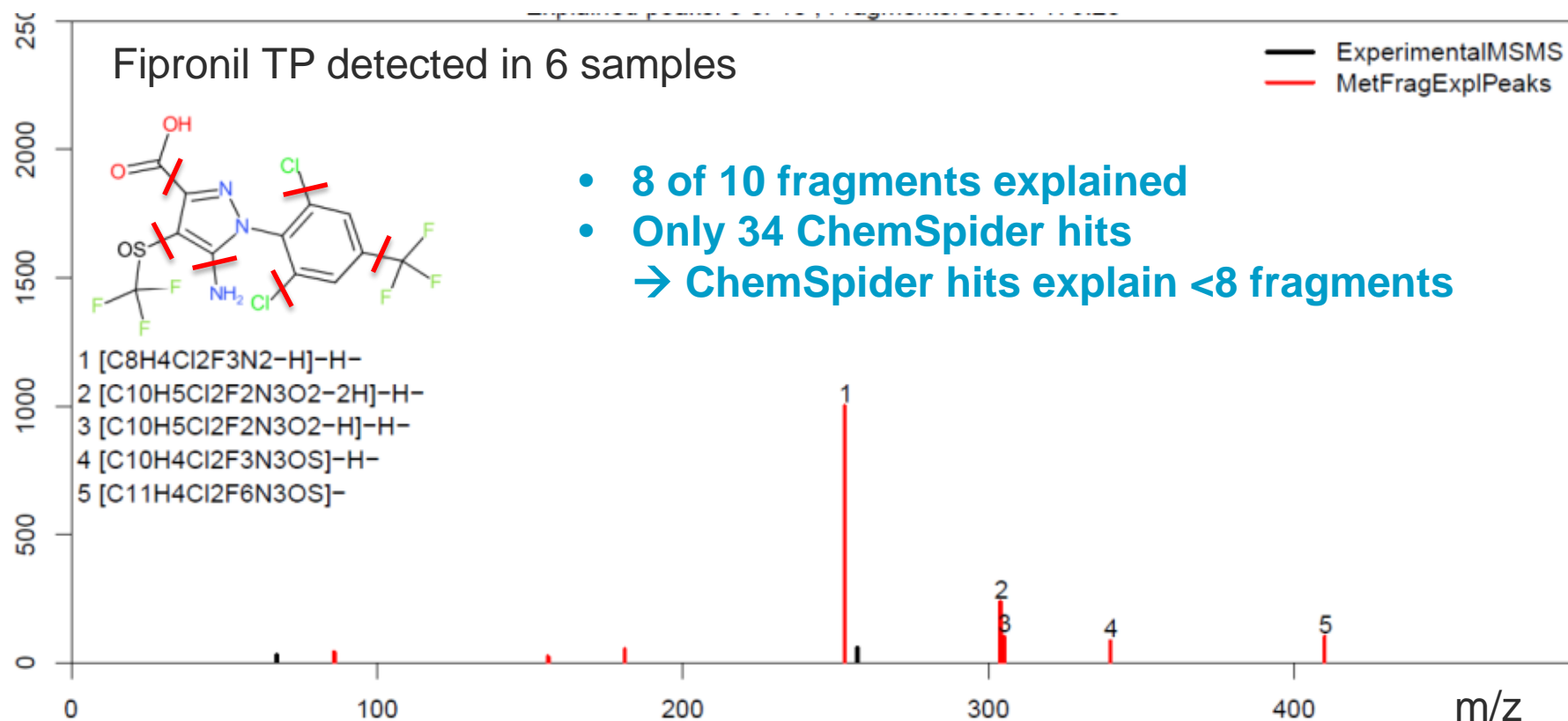
- ▶ 97% of internal standards detected
- ▶ but 9300 suspects detected
- ▶ filtering of noise/background (90%) => 686 hits
- ▶ further filtering needed



Filtering of data: in silico fragmentation with MetFrag

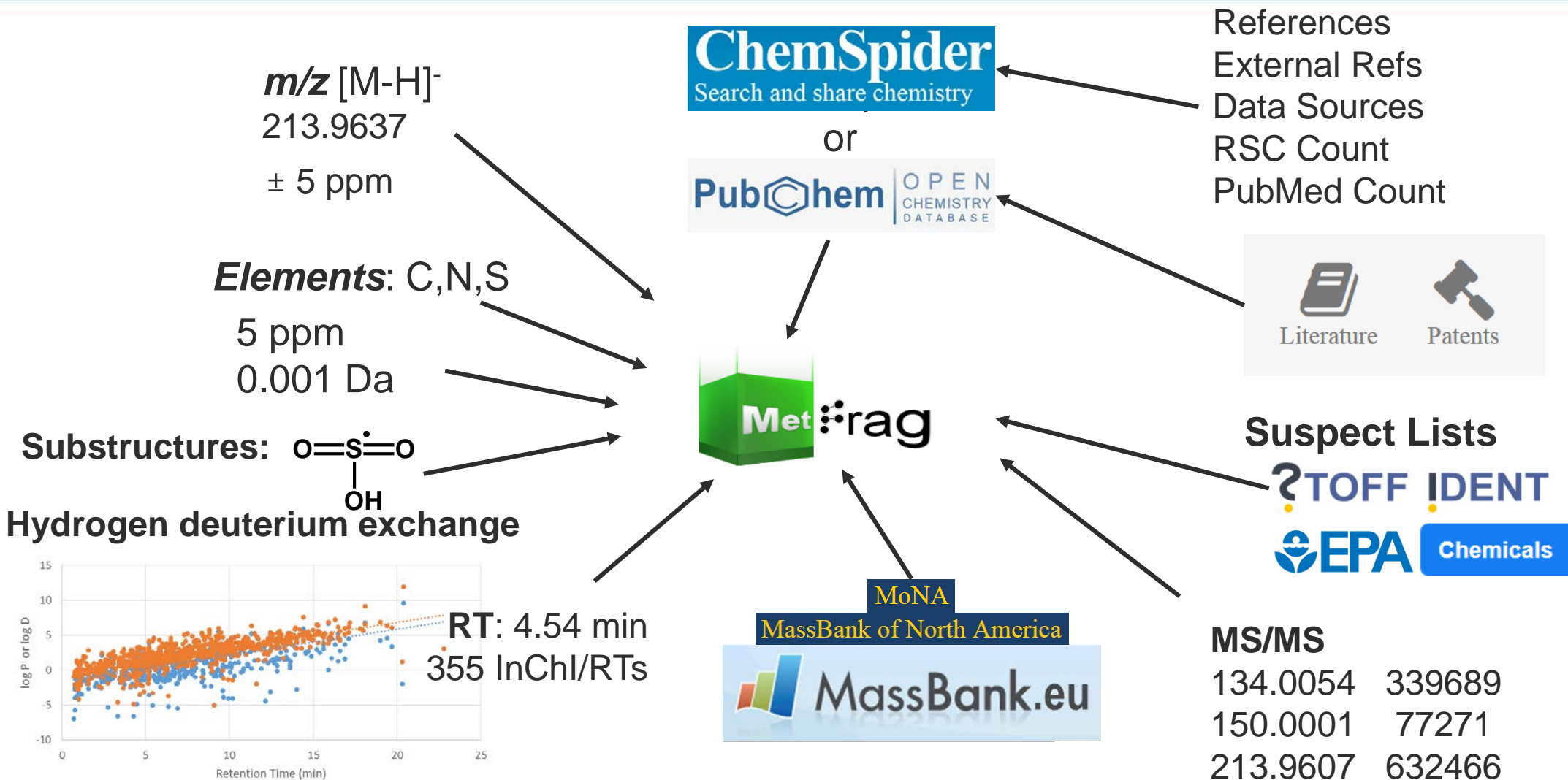


<http://c-ruttkies.github.io/MetFrag/>



- Compare against other databases
- Check also for in-source fragmentation

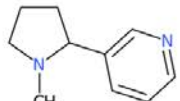
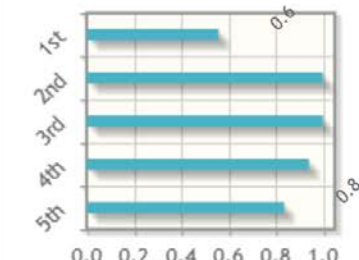
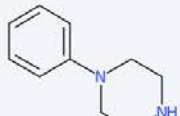
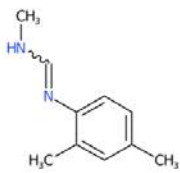
Use of meta information improves identification success



$$Score_{Final} = \omega_{Frag} * score_{Frag} + \omega_{RT} * score_{RT} + \omega_{ref} * score_{ref} + \dots$$

Ruttkies et al. *J. Cheminf.*, 2016,
Ruttkies et al., ABC 2019

Clear chemical identifiers necessary

#	Molecule	Identifier	Mass	Formula	Normalized Scores	FinalScore	Details
1	 Nicotine	<u>DTXSID1020930</u> DTXSID8021725 DTXSID3048154 DTXSID0046351 DTXSID6020931 DTXSID00657553 DTXSID5075319 InChIKeyBlock1 = SNICXCGAKADSCV	162.11576	C ₁₀ H ₁₄ N ₂		4.3349	Peaks: 18 / 23 Fragments Scores Download
2	 Phenylpiperazine	<u>DTXSID3057855</u> DTXSID4076612 DTXSID40133102 DTXSID90236632 DTXSID50293046 DTXSID00293011 DTXSID50296603 InChIKeyBlock1 = YZTJYBJCZXZGCT	162.11576	C ₁₀ H ₁₄ N ₂			
3	 N'-(2,4-Dimethylphenyl)-N-methylformamidin	<u>DTXSID1037696</u> DTXSID10199510 InChIKeyBlock1 = JIIOLEGNRQDIP	162.11576	C ₁₀ H ₁₄ N ₂			

LEGEND: Name, SMILES
DTXSID | InChIKey 1st Block
CAS | Monoiso. Mass | logP | Sources
Data on: Toxicity | Exposure | Bioassays

Nicotine
CN1CCC[C@H]1C1=CN=CC=C1
DTXSID1020930 | SNICXCGAKADSCV
54-11-5 | **162.1157** | 0.929 | **72**
Tox: **yes** | Expo: **yes** | Bioassay: **yes**

D-Nicotine
CN1CCC[C@@H]1C1=CN=CC=C1
DTXSID0046351 | SNICXCGAKADSCV
25162-00-9 | **162.1157** | 0.929 | **20**
Tox: **no** | Expo: **yes** | Bioassay: **yes**

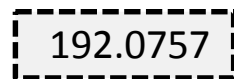
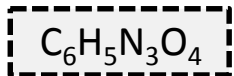
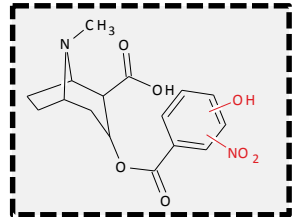
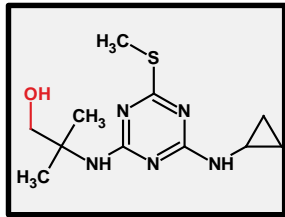
Nicotine hydrochloride
Cl.CN1CCC[C@H]1C1=CN=CC=C1
DTXSID6020931 | HDJBTCAJIMNXEW
2820-51-1 | **198.0924** | 0.929 | **9**
Tox: **no** | Expo: **yes** | Bioassay: **yes**

MS-ready
DL-Nicotine
CN1CCCC1C1=CN=CC=C1
DTXSID3048154 | SNICXCGAKADSCV
22083-74-5 | **162.1157** | 0.953 | **9**
Tox: **yes** | Expo: **no** | Bioassay: **yes**

DL-Nicotine-d3
[2H]C([2H])([2H])N1CCCC1C1=CN=CC=C1
DTXSID80442666 | SNICXCGAKADSCV
69980-24-1 | **165.1345** | 0.929 | **1**
Tox: **no** | Expo: **no** | Bioassay: **no**

“MS-ready”
form

Example



Identification confidence

Minimum data requirements

Level 1: Confirmed structure by reference standard

MS, MS², RT, Reference Std.

Level 2: Probable structure
a) by library spectrum match
b) by diagnostic evidence

MS, MS², Library MS²
MS, MS², Exp. data

Level 3: Tentative candidate(s) structure, substituent, class

MS, MS², Exp. data

Level 4: Unequivocal molecular formula

MS isotope/adduct

Level 5: Exact mass of interest

MS

What match value accepted?

How many fragments in data dependent /data independent acquisition?

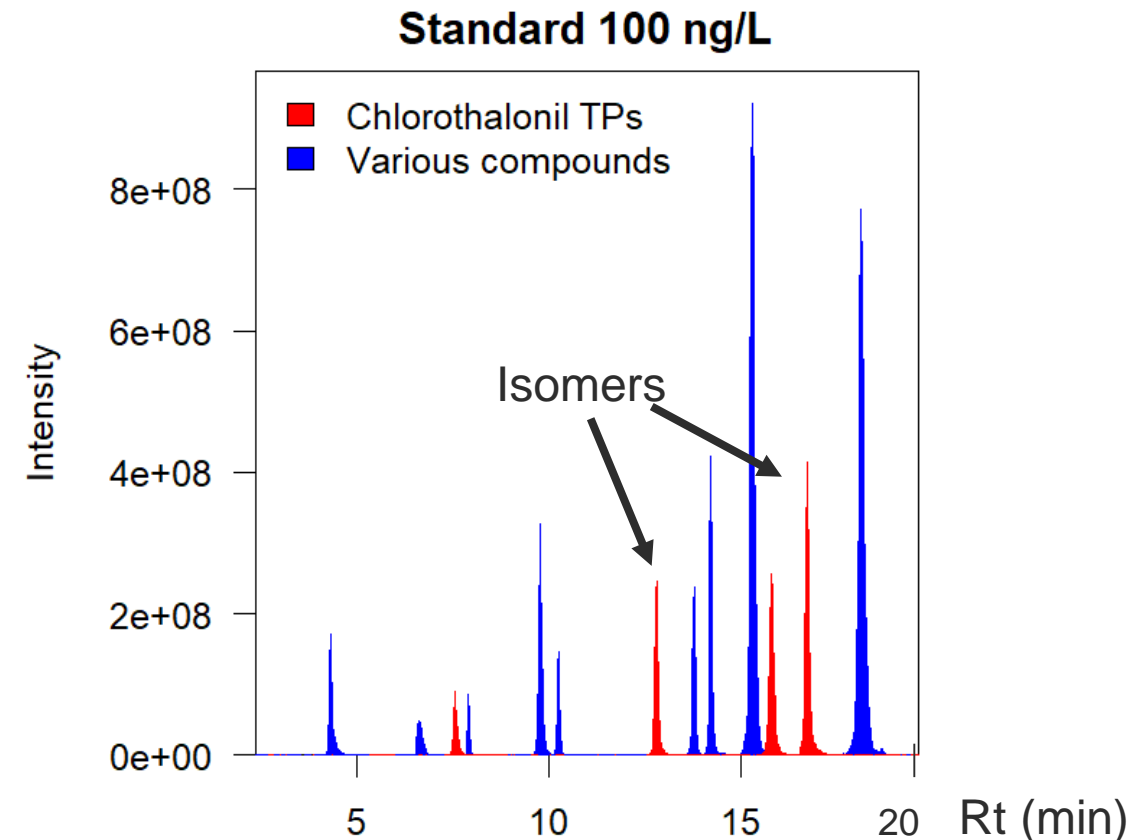
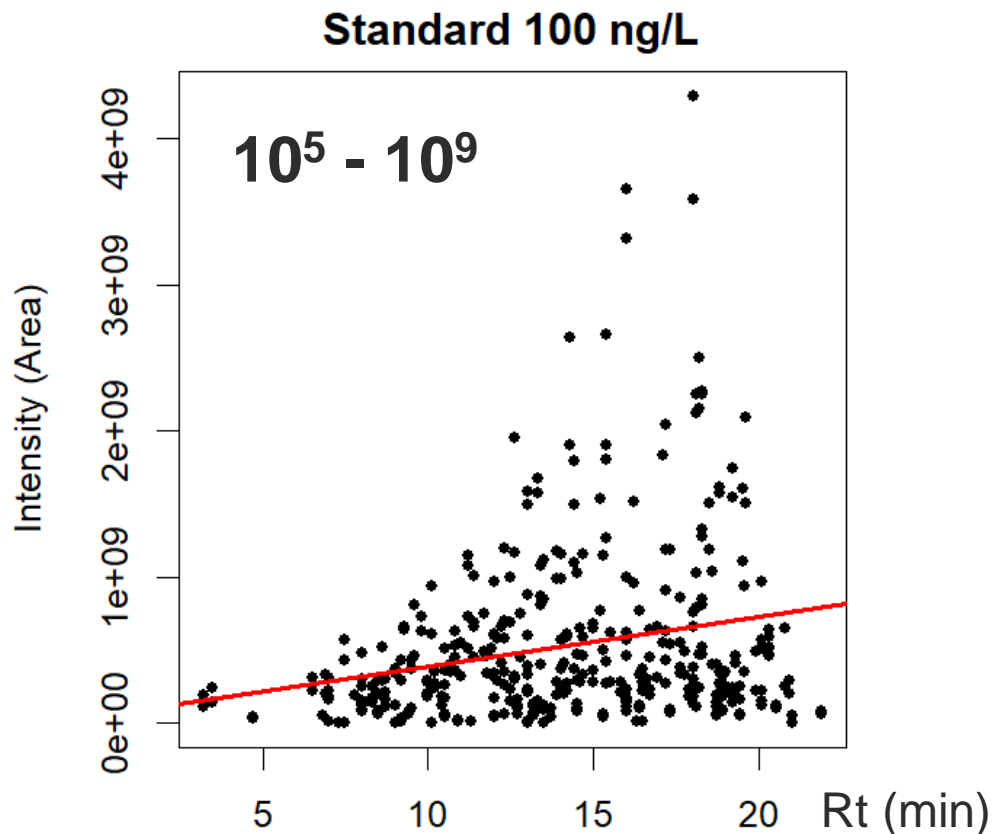
Which exp. data have how much value?

Communicating confidence of identification: Example pesticide TPs

19 Confirmed (Level 1)	<ul style="list-style-type: none">• 3 chlorothalonil TPs• 1 cycloxydim TP• 1 dimethachlor TP• 1 fipronil TP• 2 fluxapyroxad/bixafen TPs• 1 fludioxonil TP	<ul style="list-style-type: none">• 1 metalaxyl TP• 2 metolachlor TPs• 2 nicosulfuron TPs• 1 pinoxaden TP• 4 terbuthylazine TPs
2 Probable (Level 2)	<ul style="list-style-type: none">• 1 chlorotoluron TP	<ul style="list-style-type: none">• 1 fipronil TP
3 Tentative (Level 3)	<ul style="list-style-type: none">• 2 chlorothalonil TPs	<ul style="list-style-type: none">• 2 cymoxanil TPs
9 Rejected	<ul style="list-style-type: none">• 9 TPs	

(Semi) Quantification without standards

- Suspected structure**
 - with structural similar compounds, e.g. TPs with parent compound
 - Quantitative structure response relationship (e.g. Tanimoto coefficient)
- Unknown structure**
 - Concentration range based on standards with different response ratios (Hollender Wat Res X 2018)
 - Average response ratio of standards at similar retention time



- Instruments & tools are available, application in a clever way is important to successfully and reliably identify new compounds
 - Optimization of acquisition and data processing is key
 - Meta data are very important for successful identification
 - Sharing of spectra, data, standards, experiences etc. is useful (NORMAN network)
-
- Which kind of quality control samples (composites, replicates,...) and how often?
 - How to optimize identification workflow?
 - How to select smart suspect lists?
 - What kind of meta information are most useful?
Retention time, collision cross section in ion mobility, number of references, exposure index,...?
 - What are relevant differences in NTS for different samples (environment, food, human)?
 - How to improve communication of identification confidence? DDA or DIA spectra acquisition?
 - How to improve data sharing of spectra, suspect lists, etc?

Acknowledgements



Schweizerische Eidgenossenschaft
Confédération suisse
Confederazione Svizzera
Confederaziun svizra

Bundesamt für Umwelt BAFU



- Cantonal labs, Federal Institute of Metrology for groundwater sampling
- Syngenta, BASF, Dupont, Bayer for reference standards
- Steffen Ruppe, Dorit Griesshaber, Ingrid Langlois, Jan Mazacek, AUE Basel-City
- Tobias Schulze, UFZ
- Reza, Aalizadeh, University of Athens
- Nikiforos Alygizakis, Jaroslav Slobodnik, Environmental Institute
- Saer Samanipour, Kevin Thomas, NIVA
- Christoph Ruttkes, Steffen Neumann, IPB Halle



KEMI
Kemikalieinspektionen

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Non-Target Analysis for Environmental Assessment

26–30 May 2020 | Durham, NC, USA

SETAC North America Focused Topic Meeting

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A decorative background for the main content area featuring a colorful waveform. The waveform starts with purple and blue on the left, transitions through teal and green in the middle, and ends with yellow and light green on the right. The text 'Save the Date' and '26–30 May 2020' is overlaid on this waveform.

Save the Date
26–30 May 2020